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Nanoscience in Multiple Sclerosis

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Multiple Sclerosis (MS) – the most common autoimmune disease of the central nervous system – is traditionally diagnosed by methods mainly using magnetic resonance techniques to detect the lesions. Use of nanoparticles as the contrast agents can help in better diagnosis of the damaged cells. Iron nanoparticles are used in various advanced techniques like superparamagnetic iron oxide nanoparticles (SPIONs) and ultra-small SPIONs (USPIONs). The major challenge in treatment of MS is the delivery of the drug into the brain, crossing the blood brain barrier (BBB). Nanoparticles like liposomes, nanoshells, dendrimers, nanogels, micelles have potential applications in the same. Presently, no significant treatment is devoid of side effects like fever, headache and fatigue, Use of nanoscience in MS in drug delivery and treatment can help solve the prevailing inadequacies. Administered quantum dots conjugated with self-antigens act on lymph nodes and spleen. These assemblies produce regulatory T-cells which prevent degeneration of myelin sheath. New studies study modifications to produce inflammation-resistant myelin by inducing response in lymph nodes during T-cell priming. This review aims to briefly describe the application of nanotechnology in diagnosis, drug delivery and treatment of MS.

1. Introduction

Multiple sclerosis (MS) is a chronic demyelinating disease of the central nervous system. Inflammation in the central nervous system and plaque of demyelinated neurons are the major indications of MS. It is an autoimmune disease involving the innate immune system i.e. T cells and B cells. Innate immunity plays a vital role in the onset and progression of the disease. T cells have specific myelin protein targets. Presence of different T lymphocytes can indicate attack on specific myelin proteins^{1,2}. MS has affected over two million individuals and the count increases. Since its discovery in the 19th century, there has not been any significant development in the clinical management of MS until last 50 years. In the initial stages, MS diagnosed by the medical history of the patient. Recently, blood tests and lumbar puncture tests were used for the diagnosis of MS. The development of magnetic resonance imaging (MRI) greatly simplified the diagnosis of this neurodegenerative disease. Traditional MRI techniques are used for early diagnosis and estimation of its progression and the response of the cells to the treatment. After newer techniques like MR spectroscopy, magnetization transfer imaging, MS has been better studied^{3,4}. However, the diagnosis as well as treatment methods are insufficient as they are partially effective and cannot reduce the progress rate of disease.

Nanotechnology deals with materials of particle size of about 1-100 nm. Nanomedicine is the application of nanomaterials in biological systems. Its physicochemical properties offer a hope in developing new therapeutic methods in diagnosis and treatment of several diseases⁵. This review focuses on the application of nanotechnology and nanosciences in multiple sclerosis for diagnosis, drug delivery and treatment.

2. Diagnosis

Diagnosis of multiple sclerosis is difficult due to the nonspecific nature of this disease. A combination of various tests and imaging techniques are generally used for diagnosis. However, they lack sensitivity, and permeability through the blood brain barrier (BBB) and decrease half-life after administration of the diagnostic agent intravenously⁶. The ability of conventional imaging techniques to detect lesions depends upon the dosage of contrasting agent, the imaging parameters and the field strength^{7,8,9}. Nanoparticles are nanoscale solid colloidal particles composed of polymers or solid lipids. Those particles with size greater than 100 nm are used for drug delivery. The techniques generally used for diagnosis of such diseases are positron emitted tomography (PET), MRI and single photon emission computed tomography (SPECT). The use of nanoparticles might improve the neuroimaging power if better contrasts and better targeted molecular imaging probes are developed^{10,11,12,13,14,15}

The use of nanotechnology in diagnosis of the cellular inflammation is mainly seen in development of new contrast agents which have better reach and activity owing to their small size. The common markers used to detect cellular



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inflammation in MS are quantum dots, superparamagnetic iron oxide nanoparticles (SPIONs) and gadolinium-DTPA (Gd-DTPA). Gold nanoparticles (AuNPs) and Silver Nanoparticles (AgNPs) are used in the detection and amplification of molecular signals. For early detection of MS, radio diagnoses such as MRI and SPION are most common in which these contrast agents can be used to detect weak signals even at molecular concentrations. Localized Surface Plasmon Resonance (LSPR) is a cheaper and ultrasensitive technique involving Silver nanoparticles (AgNPs). In this method, the changes in the refractive index of the magnetic field due to interactions AgNPs with protein marker. Scanning tunneling microscopy-based technique can detect concentrations as low as 10fg/mL by indications in pulse-like peaks of tunnel microscope. Gold nanoparticles (AuNPs) conjugated with proteins can detect concentrations as low as 1pg/mL¹⁶.

Latest research also reveals that ultra-small superparamagnetic particles of iron oxide nanoparticles (USPION) can monitor cell infiltration more accurately over traditional methods. In-vivo techniques have already proven that USPIONs work on CD4+ lymphocytes cell mostly of midbrain and interbrain and CD8+ cells of brainstem region. These USPIONs conjugate with anti-CD4 antibodies and detect pathological regions in MS when injected intravenously in rat brain.

2.1. SPIONs

SPIONs are based on core of magnetite (Fe₃O₄) or maghemite (y-Fe₂O₃) stabilized with a hydrophilic surface coating^{17,18,19}. These nanoparticles are not only biocompatible but also show useful physicochemical properties. These particles contain only a single domain and hence exhibit superparamagnetism²⁰. Their properties enable them for their applications in MRI as contrast agents and in multimodal imaging, etc. SPIONs have been in use as contrast agents for MRI for last many years. They have been classified into Standard SPIONs (150-380 nm), very small SPIONs (< 10nm) and ultrasmall SPIONs (10-50 nm) based on the particle size²¹. SPIONs are produced by thermal decomposition of iron salts²², then are coated with surface complexing agents, and then prevented from agglomeration. SPIONs are injected intravenously to detect the lesions formed in the liver whereas USPIONs, due to their higher plasmic life, their uptake is not as fast as SPIONs, and they can reach macrophages or normal cells in affected tissues^{23,24,25,26}. Human transferrin proteins coupled with SPIONs were injected into tumour bearing rats and a 40% change in the signal intensity was observed²⁷. Another study was conducted by MRI and electron paramagnetic resonance (EPR) on SPIONs injected in the inflamed muscles. The mean iron oxide concentration in inflamed cells was 0.8% and 0.4% of the dose injected initially. When the same study was done by EPR, it was observed that concentration of the grafted USPIONs was higher by two times in the muscles than the ungrafted ones. Hence, association of SPIONs helps in enhancing the sensitivity of inflamed cell detection. SPIONs are phagocytosed by the macrophages when administered into systemic circulation. Hence, they show their paramagnetic effect by MRI in CNS disorders like MS.

3. Drug Delivery

The blood brain barrier (BBB) is a semipermeable system composed of endothelial cells of brain capillaries, astrocyte foot process and pericyte. Endothelial cells of capillaries in the blood differ from normal capillaries in presence of tight junctions. Astrocyte and pericyte ensure the transfer of essential components like glucose, amino acids including some gases, lipophilic molecules with molecular weight < 600 Dalton. Hydrophilic or larger molecules are prohibited to enter central nervous system and cerebrospinal fluid. Hence crossing the BBB becomes a major challenge in the treatment of multiple sclerosis²⁸.

Another important factor in selection of suitable drug delivery systems like liposomes is a novel drug carrier system offering a potential approach to resolve this problem.

3.1. Liposomes

Liposome is a sphere of concentric lipid bilayers with an internal aqueous cavity which traps the drug. After several years of research, liposomes have been found to have drug carrying capacities. The structural property of liposome forming barrier between the drug and the non-targeted tissue helps in effective formulation and administration of toxic drugs. Liposomes can penetrate through the blood brain barrier via various mechanisms i.e. triggered drug release, cationation of vectors, targeting ligands etc.²⁹. Liposomal complexes of doxorubicin, anthracycline derivative have been tested. Mitoxantrone, an immunosuppressant, can be a suitable candidate for use of liposomal preparations in the treatment of MS³⁰. Recently studied glucocorticoids loaded in nano-sterically stabilized liposome (nSSL) showed a therapeutic efficiency in the Proteolipid Protein (PLP) induced Experimental Autoimmune Encephalomyelitis (EAE). In animal models, recovery from acute disease was faster when treated with nSSL than with the MS drugs, Betaferon and Copaxone³¹.

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Antibodies against Myelin basic protein (MBP) are considered as cause of MS³². Mannosylation of liposomes loaded with MBP fragments decrease the rate of anti-MBP antibodies and facilitates their uptake through mannose receptors present on macrophages³³. Targeting ability of liposomes is enhanced by conjugating its surface with some ligands like glucose, lactoferrin, transferrin, specific peptides etc. Surface modified liposomes efficiently cross the BBB and are able to deliver the drug at the particular site. This advanced nanotechnological approach has proven to reduce the progress rate of MS by preventing plaque formation and removing the already formed amyloid deposits in preclinical studies³⁴.

3.2. Nanocapsules and Nanospheres

Nanocapsules and nanospheres are systems of size range 10-1000 nm³⁵. They are composed of natural nanoparticles like chitosan, alginate and/or synthetic nanoparticles like poly(lactide-co-glycolide) (PLGA), poly-lactic acid (PLA), polymethacrylic acid (PMA) and polyethylene glycol (PEG)³⁶. Nanocapsules are thin polymeric envelopes enclosing oil-filled cavities while nanospheres have solid core with a dense polymeric matrix³⁷. They are highly stable, easy to synthesize and can escape without being recognized by macrophages. In application, indomethacin-loaded nanocapsules were found to protect in-vitro hippocampal cultures against inflammation³⁸.

3.3. Dendrimers

Dendrimers are polymeric, three dimensional, monodisperse and highly branched materials which can entrap and/or conjugate with active molecules. Dendrimers show high water solubility, stability, permeability of drug^{39,40}, biocompatibility⁴¹, polyvalency⁴² and precise molecular weights⁴³, which makes them an ideal carrier for drug delivery. Various types of dendrimers such as polyamidoamine (PAMAM), poly(propylene imine) (PPI), poly-L-lysine, melamine, poly(etherhydroxylamine) (PEHAM), poly(esteramine) (PEA) and polyglycerol have been synthesized and studied as drug delivery vehicles^{44,45}. Neutral G3-G4 phosphate dendrimers are recently explored for their ability to reduce secretion of proinflammatory cytokines from mice and human monocytederived macrophages⁴⁶. To increase the activity of therapeutic agents in brain tissue, dendrimers are conjugated with CNStargeting molecules such as transferrin47,48, lactoferrin49, Dglucosamine⁵⁰ and DGL-PEG-leptin30⁵¹. In vitro and in vivo studies have demonstrated the low toxicity of dendrimers⁵². Dendrimers can act on both astrocytes and microglial cells which are involved in inflammation in diseases like MS⁶⁸.



The activation and proliferation of CD4+ T lymphocytes in Interleukin-2 (IL-2) stimulated Peripheral Blood Mononuclear Cells (PBMC) is inhibited by phosphorus-containing dendrimers with an N_3P_3 (cyclotriphosphazene) core and PMMH (phenoxymethyl-methylhydrazine) branches^{53,54}.

3.4. Micelles

Micelles are spherical arrangement of lipid molecules of 20 - 200 nm due to the amphipathic nature of lipids composed of polyethylene glycol (PEG), polypropylene glycol, etc. In normal micelles, hydrophobic tails are arranged inside and hydrophilic heads form the outer surface. Micelles may have reverse structure, i.e. hydrophobic core and hydrophilic surface, in water-in-oil conditions⁵³.

3.5. Nanogels

Nanogels are cross-linked networks which can encapsulate oligonucleotides, siRNA, DNA, proteins, and low-molecularmass drugs delivering them across the BBB. The drug-loading capacity of nanogel is up to 40–60%⁵⁴. *In vivo* studies suggested that nanogels increased brain uptake of oligonucleotides while decreasing uptake in the liver and spleen^{55,56}. Poly N-isopropylacrylamide (PNIAM) is a thermoresponsive polymer which has water retaining capacity. Hydrogel formed with PNIAM has been studied successfully in Zebrafish model for sustained drug release and compatibility. PNIAM hydrogel is a functionalized nanogel loaded with donepezil with polysorbate-80⁵⁷.

3.6. Pomegranate Seed Oil

PSO has higher levels of a unique polyunsaturated fatty acid, punicic acid. Punicic acid is a natural antioxidant. Its higher concentrations are needed for its antioxidant action. Nanodroplet formulation of PSO, labelled as nano-PSO has been tested in mice induced for EAE and gives desirable results at relatively lower concentrations reducing the oxidation of lipids and demyelination in EAE mice. An addition of other antioxidants and beta-sitosterol enhanced the activity over individual ingredients as it accumulates in plasma membrane of brain cells. Nano-PSO can be used in combination with other MS medications such as natalizumab^{58,59}.

4. Treatments

Nanoscience is a promising approach in treatment of MS⁶⁰. Nanoparticles can be readily synthesized at laboratory scale. They can carry nano medicines to the site of action by protecting



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the degeneration of the API, enhancing its bioavailability⁶¹. It has gathered attention as it involves action at the cellular and subcellular levels. Cytobots and karybots are some examples of synthetic magnetic nanomaterials which work on electrochemical principles and interact directly with the lesions in the nervous system formed during MS^{62,63}. Due to nanotechnological advancements like quantum dots used in nanoimaging, surgical tweezers, and AFM tips as sharp needle nano scissors, newer approaches like single cell surgery are also possible⁶⁴.

Nanoparticles are mainly applied in two ways – by neuroprotection or as anti-inflammatory agents, or both⁶⁵ (e.g. fullerene and its derivatives are used as neuroprotective anti-inflammatory agents). Gold NPs stabilized with a layer of PEG molecule administered in mice has been shown to attenuate disease and population expansion of regulatory T-cells.

Nano-suspensions and nanoemulsions are well-known for their target drug delivery by establishing an interaction with the cell membrane⁶⁹. Few nanoparticles can also permeate through the gaps present between the blood brain barrier by transcytosis^{70,71}. Carbon nanotubes and nanowires are being used for neural regeneration by showing cellular signal transmission⁷².

4.1. Force Tip Microscopy

The AFM tip can directly penetrate through the cell membrane into the nucleus, with the membrane returning to its original shape. This technique can deliver specific monoclonal antibodies (MAB) by coating it onto the tip which will interact with the intracellular protein traffic and track the cell in real time to study its chemistry⁶⁰.

4.2. Exosomes

Exosomes are RNA or proteins that are derived from and are created by the dendrites which help in regeneration of myelin sheath⁷³. These are similar to liposomes but enriched with adhesive molecules, cytoplasmic enzymes, signal transduction, functional mRNA microRNA. Being natural particles, they have an edge over artificial NPs. Exosomes are capable of transferring specific immunosuppressive molecules to the brain^{74,75}. Serum exosomes also increase the myelin content, oligodendrocyte-precursor cell and neural stem cell levels⁷⁶.

4.3. Cerium Oxide NPs

A new study suggests that the combination of lenalidomide and cerium oxide nanoparticles reduced demyelination⁶⁶. Cerium oxide NPs fluctuate between +3 and be +4 valence states. It has recently been shown that the intravenous administration of cerium oxide NPs into mice reduced reactive oxygen species levels and disease attenuation. Thus, cerium oxide nanoparticles may be effective in the MS therapy in part through reduction of oxidative stress process⁶⁷, which is the primary observation in any neurodegenerative disease. They either receive or donate an electron and hence reduce ROS and enhance brain activity^{77,78}.

4.4. Nanocurcumin

Curcumin has been studied for its pharmacological activities like its anti-inflammatory, anti-tumor, antifungal, antibacterial, antioxidant and antiprotozoal activity79,80,81. Even though curcumin has a large variety of therapeutic activities, its use was limited due to its poor solubility and bioavailability. Addition of dendrosome nanoparticles unfolds its application. Polymerised nano curcumin (PNC) has been studied in EAE model; it inhibits neuroinflammation by blocking cytokines pathway. Initial PNC administration has reduced development of EAE score while daily dose decreased the relapsing symptoms. According to recent studies, nanocurcumin can be a massive treatment to inhibit disease progression in MS patients^{82,83,84}. Curcumin is a polyphenolic non-enzymatic antioxidant which removes free radicals⁸². Its use is limited due to insolubility in body fluids, low intrinsic activity and rapid clearance from the body and overall low bioavailability. To overcome these problems, a number of solutions have been examined by adjuvants, liposomes and other NPs⁸⁵.

5. Conclusion

In the last few decades, a lot of new studies have been conducted to find out new potential methods for drug delivery in MS. The applications of nanotechnology in MS have resolved the traditional problems, and promise better imaging, drug delivery and treatment. Use of these nanomaterials will not only improve the efficiency of the therapeutic applications but also provide an easy, cost-efficient and potentially a widely accepted method for the same. These techniques can track a particular disease at the cellular or molecular level. Nanosurgery and nano neuroprotection have gained a lot of attention due to some promising applications. However, there are still some challenges to fully study the implications of nanomaterials and a lot of research on their application in disease diagnosis and treatment has to be done. Their safety should also be evaluated.



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6. Future Scope

There have been a lot of developments in drug delivery and treatment of MS in last few years. Many new drugs have been developed and approved in diagnosis and treatment of MS but still a lot needs to be done. The use of nanoscience has greatly influenced the methods by which we can treat MS and in upcoming years with more advancement we can develop better models. The development of nanodrug carriers has allowed better targeted action of the drug, site specificity, real time monitoring, effective action and higher sensitivity, which are some advantages that we have already achieved. There are still many limitations on the application of nanotechnology in neuroscience and one must also study the potential hazards of the administration and safety of nanomedicines. The upcoming decade will hold a lot of progress in nanotechnology and its applications is a wide range of diseases.

7. Keywords

Multiple sclerosis, nanoparticles, blood brain barrier, quantum dots.

8. References

- B. Bielekova, B. Goodwin, N. Richert, I. Cortese, T.Kondo, G. Afshar et al., Encephalitogenic potential of the myelin basic protein peptide (amino acids 83-99) in multiple sclerosis: results of a phase II clinical trial with an altered peptide ligand, Nat Med. 6(2000), 1167-75. https://doi.org/10.1038/80516
- E.M. Chastain, D.S. Duncan, J.M. Rodgers, S.D. Miller, The role of antigen presenting cells in multiple sclerosis, Biochim Biophys Acta, 1821 (2011), 265-274. https://doi.org/10.1016/j.bbadis.2010.07.008
- McDonald, W. I., Compston, A., Edan, G., Goodkin, D., Hartung, H. P., and Lublin, F. D., 2001. Recommended diagnostic criteria for multiple sclerosis: guidelines from the International Panel on the Diagnosis of Multiple Sclerosis. Ann. Neurol. 50, 121–127. https://doi.org/10.1002/ana.1032
- Polman, C. H., Reingold, S. C., Edan, G., Filippi, M., Hartung, H. P., and Kappos, L., 2005. Diagnostic criteria for multiple sclerosis: 2005 revisions to the "McDonald Criteria". Ann. Neurol. 58, 840–846. https://doi.org/10.1002/ana.20703
- M. Mahmoudi, V. Serpooshan, S. Laurent, Engineered nanoparticles for biomolecular imaging, Nanoscale. 3 (8), 3007-3026.

- A.V. Singh, R. Patil, D.K. Thombre, W.N. Gade, Micronanopatterning as tool to study the role of physicochemical properties on cell-surface interactions, J Biomed Matter Res A. 10(2013) 3019-3032. doi: 10.1002/jbm.a.34586
- Keiper MD, Grossman RI, Hirsch JA, et al., 1998. MR identification of white matter abnormalities in multiple sclerosis: a comparison between 1.5T and 4T. AJNR Am J Neuroradiol.19:1489–93.
- Wolansky LJ, Bardini JA, Cook SD, et al.,1994.Triple-dose versus single-dose gadoteridol in multiple sclerosis patients, J Neuroimaging. 4:141–45.
- Sardanelli F, Iozzelli A, Losacco C, et al., 2003. Three subsequent single doses of gadolinium chelate for brain MR imaging in multiple sclerosis. AJNR Am J Neuroradiol. 24:658–62.
- K. Tucker, K.R. Robertson, W. Lin, J.K. Smith, H. An, Y. Chen et al., Neuroimaging in human immunodeficiency virus infection, J Neuroimmunol, 157 (2004),153–62. doi: 10.1016/j.jneuroim.2004.08.036
- C. Bocti, Topographical patterns of lobar atrophy in frontotemporal dementia and Alzheimer's disease, Dement Geriatr Cogn Disord, 21 (2006), 364–72. doi: 10.1159/000091838
- D.H. Silverman, A. Alavi, PET imaging in the assessment of normal and impaired cognitive function, Radiol Clin North Am, 43 (2005),67–77.
- S.L. Wearne, New techniques for imaging, digitization and analysis of three-dimensional neural morphology on multiple scales, Neuroscience, 136 (2005), 661–80. https://doi.org/10.1016/j.neuroscience.2005.05.053
- G.A. Silva, Neuroscience nanotechnology: progress, opportunities and challenges, Nat. Rev. Neurosci, 7 (2006), 65-74. doi: 10.1038/nrn1827
- D.J. Burn, J.T. O'Brien, Use of functional imaging in Parkinsonism and dementia, Mov Disord, 18 (2003), S88– 95. https://doi.org/10.1002/mds.10568]
- A.V. Singh, M. Khare, W. N. Gade, P. Zamboni, Theranostic Implications of Nanotechnology in Multiple Sclerosis: A Future Perspective, Autoimmune Diseases, 160830 (2012). doi:10.1155/2012/160830
- Laurent, S., Forge, D., Port, M., Roch, A., Robic, C., Vander Elst, L., and Muller, R. N. (2008) Magnetic iron oxide nanoparticles: Synthesis, stabilization, vectorization,physicochemical characterizations and biological applications. Chem. Rev. 108, 2064–2110.
- Mahmoudi, M., Sant, S., Wang, B., Laurent, S., Sen, T. (2011) Superparamagnetic iron oxide nanoparticles (SPIONs): Development, surface modification and applications in chemotherapy, Adv. Drug Delivery Rev. In



ARTICLE

press. https://doi.org/10.1016/j.addr.2010.05.006. [19] Laurent, S., Bridot, J. L., Vander Elst, L., and Muller, R. (2010) Magnetic iron oxide nanoparticles for biomedical applications. Future Med. Chem. 2, 427–449.

- Mahmoudi, M., Hosseinkhani, M., Boutry, S., Simchi, A., Hosseinkhani, H., Journeay, W. S., Subramani, K.,Laurent, S. (2011) MRI tracking of stem cells in vivo using iron oxide nanoparticles as a tool for the advancement of clinical regenerative medicine,Chem. Rev.In press. doi: 10.1021/cr1001832.
- Corot, C., Robert, P., Idee, J. M., and Port, M. (2006), Recent advances in iron oxide nanocrystal technology for medical imaging. Adv. Drug Delivery Rev. 58, 1471–1504.
- Mahmoudi, M., Milani, A. S., and Stroeve, P. (2010) Surface Architecture of Superparamagnetic Iron Oxide Nanoparticles for Application in Drug Delivery and Their Biological Response: A Review. Int. J. Biomed. Nanosci. Nanotechnol. 1, 164–201.
- Vellinga, M. M., Vrenken, H., Hulst, H. E., Polman, C. H., Uitdehaag, B. M., Pouwels, P. J., Barkhof, F., and Geurts, J. J. (2009) Use of ultrasmall superparamagnetic particles of iron oxide (USPIO)-enhanced MRI to demonstrate diffuse inflammation in the normal-appearing white matter (NAWM) of multiple sclerosis (MS) patients: an exploratory study. J. Magn. Reson. Imaging 29, 774–779.
- 23. Stoll, G., and Bendszus, M. (2010) New approaches to neuroimaging of central nervous system inflammation. Curr. Opin. Neurol. 23, 282–286.
- Bonnemain, B. (2008) Nanoparticles: the industrial viewpoint. Applications in diagnostic imaging. Ann. Pharm. Fr. 66, 263–267.
- Tang, T. Y., Howarth, S. P. S., Miller, S. R., Graves, M. J., U-King-Im, J. M., Li, Z. Y., Walsh, S. R., Patterson, A. J., Kirkpatrick, P. J., Warburton, E. A., Varty, K., Gaunt, M. E., and Gillard, J. H. (2008) Correlation of carotid atheromatous plaque inflammation using USPIO-enhanced MR imaging with degree of luminal Stenosis. Stroke 39, 2144–2147.
- Kresse, M., Wagner, S., Pfefferer, D., Lawaczeck, R., Elste, V., and Semmler, W. (1998) Targeting of ultrasmall superparamagnetic iron oxide (USPIO) particles to tumor cells in vivo by using transferrin receptor pathways. Magn. Reson. Med. 40, 236–242.
- Ballabh, Praveen; Braun, Alex; Nedergaard, Maiken (2004). The blood-brain barrier: an overview: Structure, regulation, and clinical implications, Neurobiology of Disease. 16 (1): 1–13 https://doi.org/10.1016/j.nbd.2003.12.016

- D.B Vieira, L. F Gamarra, Getting into the brain: liposomebased strategies for effective drug delivery across the blood-brain barrier, Int J Nanomedicine. 11 (2016)5381– 5414. https://doi.org/10.2147/IJN.S117210
- 29. G. Batist, et al., Myocet, liposome-encapsulated doxorubicin citrate: a new approach in breast cancer therapy, Exp. Opin. Pharmacother, 3 12 (2002) 1739–1751.
- 30. Y. Avnir, K. Turjeman, D. Tulchinsky, A. Sigal, P.. Kizelsztein, D. Tzemach, A. Gabizon, Y.Barenholz, Fabrication Principles and Their Contribution to the Superior In Vivo Therapeutic Efficacy of Nano-Liposomes Remote Loaded with Glucocorticoids https://doi.org/10.1371/journal.pone.0025721
- Berger T, Rubner P, Schautzer F, Egg R, Ulmer H, Mayringer I, Dilitz E, Deisenhammer F, Reindl M (2003). Antimyelin antibodies as a predictor of clinically definite multiple sclerosis after a first demyelinating event. N. Engl. J. Med. 349,2. 139-45. https://doi.org/10.1056/NEJMoa022328
- Belogurov AA, Jr, Stepanov AV, Smirnov IV, Melamed D, Bacon A, Mamedov AE, et al. Liposome-encapsulated peptides protect against experimental allergic encephalitis. FASEB J 27 (2013), 222–231. doi: 10.1096/fj.12-213975
- 33. Agrawal M, Ajazuddin, Tripathi DK, Saraf S,Antimisiaris SG, Mourtas S, Hammarlund-Udenaes , Alexander A,.Recent advancements in liposomes targeting strategies to cross blood-brain barrier (BBB) for the treatment of Alzheimer's disease.J Control Release. 260 (2017) 61-77. https://doi.org/10.1016/j.jconrel.2017.05.019
- Müller RH, Keck CM, Drug delivery to the brainrealization by novel drug carriers. J. Nanosci. Nanotechnol. 4(2004), 471–483.
- A. L. Armstead, B. Li,Nanomedicine as an emerging approach against intracellular pathogens,International Journal of Nanomedicine 6(2011),3281-3293. doi: 10.2147/IJN.S27285
- G. Modi, V. Pillay, Y. E. Choonara, Advances in the treatment of neurodegenerative disorders employing nanotechnology, Ann N Y Acad Sci. 1184 (2010), 154-172. doi: 10.1111/j.1749-6632.2009.05108.x.
- 37. Bernardi A, Frozza RL, Horn AP, Campos MM, Calixto JB, Salbego C, Pohlmann AR, Guterres SS, Battastini AM..2010. Protective effects of indomethacin-loaded nanocapsules against oxygen-glucose deprivation in organotypic hippocampal slice cultures: involvement of neuroinflammation. Neurochem Int. 57(6):629-3610. https://doi.org/1016/j.neuint.2010.07.012.



ARTICLE

- Wong H. L.; Wu X. Y.; Bendayan R., 2012. Nanotechnological advances for the delivery of CNS therapeutics. Adv. Drug Delivery Rev. 64, 686–700.
- Soto-Castro D, Cruz-Morales JA, Ramírez Apan MT, Guadarrama P.,2012. Solubilization and anticanceractivity enhancement of Methotrexate by novel dendrimeric nanodevices synthesized in one-step reaction. Bioorg Chem.,41-2,13–21.
- 40. Duncan R, Izzo L.,2005. Dendrimer biocompatibility and toxicity. Adv Drug Deliv Rev.,;57,2215–37
- Patton DL, Cosgrove Sweeney YT, McCarthy TD, Hillier SL.,2006. Preclinical safety and efficacy assessments of dendrimer-based (SPL7013) microbicide gel formulations in a nonhuman primate model. Antimicrob Agents Chemother.,50, 1696–700.
- 42. Tomalia DA.,2005, Birth of a new macromolecular architecture: Dendrimers as quantized building blocks for nanoscale synthetic polymer chemistry. Prog Polym Sci.,30, 294–324.
- Dufès C, Uchegbu IF, Schätzlein AG., 2005, Dendrimers in gene delivery. Adv Drug Deliv Rev.,57, 2177–2202.
- Wolinsky JB, Grinstaff MW.,2008. Therapeutic and diagnostic applications of dendrimers for cancer treatment. Adv Drug Deliv Rev., 60, 1037–1055.
- 45. Posadas I, Romero-Castillo L, El Brahmi N, Manzanares D, Mignani S, Majoral JP, Ceña V, 2017 Neutral high-generation phosphorus dendrimers inhibit macrophage-mediated inflammatory response in vitro and in vivo.Proc Natl Acad Sci U S A.114(37), E7660-E7669, https://doi.org/ 10.1073/pnas.1704858114
- Jackson AL, Linsley PS. 2010. Recognizing and avoiding siRNA off-target effects for target identification and therapeutic application. Nat Rev Drug Discov. 9,57–67.
- Beg S, Samad AI, Alam M, Nazish I. 2011. Dendrimers as novel systems for delivery of neuropharmaceuticals to the brain. CNS Neurol Disord Drug Targets. 10. 576–588.
- Huang R, Ke W, Liu Y, Jiang C, Pei Y., 2008. The use of lactoferrin as a ligand for targeting the polyamidoaminebased gene delivery system to the brain. Biomaterials. 29,238–246.
- Dhanikula RS, Argaw A, Bouchard J-F, Hildgen P. 2008. Methotrexate loaded polyether-copolyester dendrimers for the treatment of gliomas: Enhanced efficacy and intratumoral transport capability. Mol Pharm., 5.105–116.
- Liu Y, Li J, Shao K, Huang R, Ye L, Lou J, Jiang C. 2010. A leptin derived 30- amino-acid peptide modified pegylated poly-L-lysine dendrigraft for brain targeted gene delivery. Biomaterials, 31. 5246–5257

- Hemmer R, Hall A, Spaulding R, Rossow B, Hester M, Caroway M, Haskamp A, Wall S, Bullen HA, Morris C. 2013. Analysis of biotinylated generation 4 poly (amidoamine)(PAMAM) dendrimer distribution in the rat brain and toxicity in a cellular model of the blood-brain barrier. Molecules, 18. 11537–11552
- Damien P., Mary P., Olivier R., Cédric-Olivier T., Jean-Jacques F., Jean-Pierre M., Anne-Marie C.,Remy P. (2001),Regulatory activity of azabisphosphonate-capped dendrimers on human CD4+ T cell proliferation enhances ex-vivo expansion of NK cells from PBMCs for immunotherapy,J Transl Med. 7: 82. https://doi.org/10.1186/1479-5876-7-82
- 53. Laurent G, Mary P., Patrice M., Alexandrine M., Cdric-Olivier T., Olivier R., Pascal M., Grard B., Jean-Jacques F., Anne-Marie C.,Rmy P., and Jean-Pierre M.(2007) ,Multiplication of Human Natural Killer Cells by Nanosized Phosphonate-Capped Dendrimers,Angewandte Chemie
- Lu C-T, Zhao Y-Z, Wong HL, Cai J, Peng L, Tian X-Q.(2014). Current approaches to enhance CNS delivery of drugs across the brain barriers. Int J Nanomedicine. 9, 2241–2257.4] Vinogradov, S.V., A.D. Zeman, E.V. Batrakova & A.V. Kabanov. 2005. Polyplex nanogel formulations for drug delivery of cytotoxic nucleoside analogs. J. Control. Rel. 107: 143–157.
- Kumar, R.M., M. Sameti, et al . 2003. Polymeric nanoparticles for drug and gene delivery. In Encyclopedia of Nanoscience & Nanotechnology. Nalwa, H.S., Ed.: 1– 19.
- Friedrich, I., S. Reichl & C.C. Muller-Goymann. 2005. Drug release and permeation studies of nanosuspensions based on solidified reverse micellar solutions (SRMS). Int. J. Pharm. 305, 167–175.
- 57. Sivaji K.,Pitchai A., Soundarapandian N., Joseph B.Appadurai M.,Samuel G., Prakash V.,Rajaretinam R.K. (2016), Development of biocompatible nanogel for sustained drug release by overcoming the blood brain barrier in zebrafish model,Jr appl Biomed, 14, 157-169. https://doi.org/10.1016/j.jab.2016.01.004
- C. Shi, et al., (2013). Incorporation of b-sitosterol into the membrane increases resistance to oxidative stress and lipid peroxidation via estrogen receptor-mediated PI3 K/GSK3b signaling, Biochimica Biophys. Acta (BBA)-Gener. 1830 (3) 2538–2544.
- Treatment of a multiple sclerosis animal model by a novel nanodrop formulation of a natural antioxidant,(2015). Binyamin O., Larush L., Frid K., Keller G., Friedman-Levi Y., Ovadia H., Abramsky O., Magdassi S., Gabizon R., Int



ARTICLE

J Nanomedicine. 10:7165-74. doi: 10.2147/IJN.S92704.eCollection 2015.

- Ajay V. S., Manish K. , W. N. Gade, and Paolo Zamboni, Theranostic Implications of Nanotechnology in Multiple Sclerosis: A Future Perspective, 2012. doi: 10.1155/2012/160830
- Smriti Ojha, Babita Kumar, A review on nanotechnology based innovations in diagnosis and treatment of multiple sclerosis, (2017) Journal of Cellular Immunotherapy. https://doi.org/ 10.1016/j.jocit.2017.12.001
- A. M. Khawaja, The legacy of nanotechnology: revolution and prospects in neurosurgery, International Journal of Surgery, 9 (2011), 608–614
- R. A. Freitas, Nanotechnology, nanomedicine and nanosurgery, International Journal of Surgery, vol. 3 (2005), 243–246.
- 64. G. D. M. Jeffries, J. S. Edgar, Z. Yiqiong, J. P. Shelby, F.Christine, and D. T. Chiu, "Using polarization-shaped optical vortex traps for single-cell nanosurgery," Nano Letters, vol. 7,no. 2, pp.415–420, 2007.
- 65. Merodio M, Irache JM, Eclancher F, Mirshahi M, Villarroya H. 2000.Distribution of albumin nanoparticles in animals induced with the experimental allergic encephalomyelitis. J Drug Target. 8:289–303.
- 66. Eitan E, Hutchison ER, Greig NH, Tweedie D, Celik H, Ghosh S, Fishbein KW, Spencer RG, Sasaki CY, Ghosh P. 2015. Combination therapy with lenalidomide and nanoceria ameliorates CNS autoimmunity. Exp Neurol. 273:151–160.
- Heckman KL, Decoteau W, Estevez A, Reed KJ, Costanzo W, Sanford D, Leiter JC, Clauss J, Knapp K, Gomez C. 2013. Custom cerium oxide nanoparticles protect against a free radical mediated autoimmune degenerative disease in the brain. ACS Nano. 7:10582–10596.
- 68. Dai H, Navath RS, Balakrishnan B, Guru BR, Mishra MK, Romero R, Kannan RM, Kannan S. 2010. Intrinsic targeting of inflammatory cells in the brain by polyamidoamine dendrimers upon subarachnoid administration. Nanomedicine. 5:1317–1329.
- 69. A.V. Singh, Recent Trends in Nano-Biotechnology Reinforcing Contemporary Pharmaceutical Design, Curr Pharm Des, 22 (2016), 1415-1427.
- 70. S.S. Ali, J.I. Hardt, K.L. Quick, J.S. Kim-Han, B.F. Erlanger, T.T. Huang et al., A biologically effective fullerene (C60) derivative with superoxide dismutase mimetic properties, Free Radic Biol Med, 37(2004),1191– 1.
- 71. J.J. Liu, C.Y. Wang, J.G. Wang, H.J. Ruan, C.Y. Fan, Peripheral nerve regeneration using composite poly(lactic

acid-caprolactone)/nerve growth factor conduits prepared by coaxial electrospinning, J Biomed Mater Res A, 96 (2011), 13–20.

- J.J. Liu, C.Y. Wang, J.G. Wang, H.J. Ruan, C.Y. Fan, Peripheral nerve regeneration using composite poly(lactic acid-caprolactone)/nerve growth factor conduits prepared by coaxial electrospinning, J Biomed Mater Res A, 96 (2011), 13–20.
- P.J. Darlington, et al., Diminished Th17 (not Th1) responses underlie multiple sclerosis disease abrogation after hematopoietic stem cell transplantation, Ann. Neurol. 73 (3) (2013) 341–354
- C. Yang, P.D. Robbins, Immunosuppressive exosomes: a new approach for treating arthritis, Int. J. Rheumatol. 2012 (2012).
- 75. W. Yin, et al., Immature dendritic cell-derived exosomes: a promise subcellular vaccine for autoimmunity. Inflammation, 36 (1) (2013), 232–240.
- 76. A.D. Pusic, R.P. Kraig, Youth and environmental enrichment generate serum exosomes containing miR-219 that promote CNS myelination, Glia 62 (2) (2014) 284– 299.
- 77. K.L. Heckman, et al., Custom cerium oxide nanoparticles protect against a free radical mediated autoimmune degenerative disease in the brain, ACS Nano 7 (12) (2013) 10582–10596.
- J.-G. Leu, et al., The effects of gold nanoparticles in wound healing with antioxidant epigallocatechin gallate and alipoic acid, (2012). Nanomed.: Nanotechnol. Biol. Med. 8 (5) 767–775.
- 79. Bengmark S, Mesa MD, Gil A. Plant-derived health: the effects of turmeric and curcuminoids, (2009). Nutr Hosp, 24, 273–81.
- Kidd PM. Bioavailability and activity of phytosome complexes from botanical polyphenols: the silymarin, curcumin, green tea, and grape seed extracts,(2009). Altern Med Rev, 14, 226–46.
- 81. Ray B, Lahiri DK. Neuroinflammation in Alzheimer's disease: different molecular targets and potential therapeutic agents including curcumin,(2009). Curr Opin Pharmacol, 9, 434–44.
- Lin Xie , Xiao-Kang Li, Shiro Takahara, Curcumin has bright prospects for the treatment of multiple sclerosis, (2011). Int Immunopharmacol, 11(3):323-30. doi: 10.1016/j.intimp.2010.08.013
- Mohajeri M., Sadeghizadeh M., Najafi F., Javan M., (2015) Polymerized nano-curcumin attenuates neurological symptoms in EAE model of multiple sclerosis through down regulation of inflammatory and oxidative processes

ARTICLE

and enhancing neuroprotection and myelin repair. Neuropharmacology,99,156-167.

https://doi.org/10.1016/j.neuropharm.2015.07.013

- Anand P, Kunnumakkara AB, Newman RA, Aggarwal (2007) Bioavailability of curcumin: problems and promises. Molecular pharmaceutics, 4(6), 807-818.
- A.M. Alizadeh, et al., Encapsulation of curcumin in diblock copolymer micelles for cancer therapy, (2015). BioMed Res. Int. 824746. doi: 10.1155/2015/824746

